



**GV-SOLAS**

Gesellschaft für Versuchstierkunde  
Society for Laboratory Animal Science

# **Expert Information**

**from the Working Group on Hygiene**

**Implication of infectious agents on  
results of animal experiments**

***Citrobacter rodentium***

**Status January 2019**

**Authors: GV-SOLAS Working Group on Hygiene**

## Contents

Background .....	3
Prevalence .....	3
Host species.....	3
Properties.....	3
Susceptibility .....	3
Organotropism.....	4
Clinical disease .....	4
Pathology .....	4
Morbidity and mortality .....	6
Zoonotic potential .....	6
Interference with research .....	6
<i>Oncology</i> .....	6
<i>Teratology</i> .....	6
<i>Infectiology</i> .....	6
<i>Immunology</i> .....	7
<i>Interactions with other infectious agents</i> .....	7
<i>Toxicology</i> .....	8
<i>Physiology</i> .....	8
<i>Cell biology</i> .....	8
<i>Assisted reproductive technology</i> .....	8
Special considerations.....	8
References.....	9

## ***Citrobacter rodentium***

### **Background**

- First isolated by Barthold et al. from an outbreak of mouse diarrhea in 1972<sup>1</sup>
- Formerly classified as *Citrobacter freundii* biotype 4280<sup>2</sup> and *Citrobacter genomospecies* 9<sup>3</sup>
- Later described by Schauer and Falkow in 1993<sup>4</sup> and renamed *Citrobacter (C.) rodentium* sp. nov.<sup>5</sup>

### **Prevalence**

- Recent screening of 82.337 samples revealed no prevalence of *C. rodentium* in laboratory mice in North America and Europe.<sup>6</sup>
- Prevalence in wild mice is unknown.<sup>7</sup>

### **Host species**

- *C. rodentium* is a natural murine intestinal pathogen.<sup>2,8,9</sup>

### **Properties**

- *C. rodentium* is transmitted via the faecal-oral route and causes colitis.<sup>10</sup>
- *C. rodentium* is not highly contagious.<sup>7</sup>
- Bacteria that are shed at the peak of infection are hyperinfectious and are efficiently spread via coprophagy to uninfected littermates.<sup>10</sup>
- *C. rodentium* possesses a genetic pathogenicity island (LEE) and virulence factors that cause attaching and effacing (AE) lesions that are responsible for the pathogenesis of transmissible murine colonic hyperplasia (TMCH).<sup>4,7,10-13</sup>

### **Susceptibility**

- *C. rodentium* appears to be highly species-specific: it is frequently associated with disease, when present in a mouse population.<sup>7</sup>
- Suckling mice, adult mice of some inbred strains<sup>8,14,15</sup>, Han:NMRI mice<sup>16</sup> and transgenic lines<sup>17</sup> are more susceptible and demonstrate clinical signs.
- Germfree CF1 and C3H mice are highly susceptible, germfree C57BL/6 and NC mice are susceptible, and germfree BALB/C are resistant to infection.<sup>1,4</sup>
- Immunodeficient mice (mice lacking IL-12, IFN- $\gamma$ , TNF receptor) and mice with deficiencies in cell-mediated immunity are more susceptible to infection than immunocompetent mice and infection is often fatal in immunodeficient mice.<sup>18-23</sup>
- Intestinal mucin (Muc2) deficient mice are highly susceptible to infection because of the disruption of the intestinal mucus layer.<sup>24</sup>
- Composition of the intestinal microbiota is an important factor that influences susceptibility to *C. rodentium* infection and the subsequent immune response.<sup>10</sup>
- Several nutrients like vitamin D, polyunsaturated fatty acids as well as deficiencies in selenium and vitamin E increase susceptibility to *C. rodentium* colonization.<sup>10</sup>
- One report about an epidemic outbreak in a gerbil colony<sup>25</sup>
- Rats and hamsters are not susceptible to infection.<sup>8</sup>

## Organotropism

- *C. rodentium* is the etiologic agent of transmissible murine colonic hyperplasia (TMCH) in mice: descending colon is most commonly affected, but the entire colon and caecum may be involved.<sup>2,26</sup>
- *C. rodentium* primary colonizes the caecum, specifically within the caecal patch; following colonization of the caecum, *C. rodentium* establishes a colonic infection.<sup>27</sup>
- Clearance of *C. rodentium* in the faeces occurs 2-3 weeks post infection.<sup>10</sup>

## Clinical disease

- Transient infection with no carrier state in immunocompetent mice lasting about 4 weeks<sup>7,13</sup>
- *C. rodentium* is often absent when clinical signs are apparent.<sup>7</sup>
- Clinical signs are nonspecific and include ruffled coat, weight loss, depression, stunting, perianal faecal staining, pasty dark faeces that smear the cage wall.<sup>7,28</sup>
- Rectal prolapse is frequently associated with TMCH.<sup>7</sup>
- Variable incidence of rectal prolapse in mice of all ages is indicative of infection.<sup>28</sup>
- Disease severity ranges from self-limiting colitis and immunity (that prevents an infection) to severe inflammation and potentially lethal dehydration.<sup>10</sup>
- Mice that recover are refractory to reinfection.<sup>13</sup>
- Wild-type mice clear the infection: T and/or B lymphocytes are required to clear the infection.<sup>20</sup>
- Survival and clearance of infection does not depend on secretory IgA or IgM antibodies, but passive immunization with serum IgG protects CD4-deficient mice from fatal infection.<sup>23</sup>
- Streptomycin in the drinking water may influence the severity of the disease.<sup>29</sup>
- *C. rodentium* has the ability to persist in mice following kanamycin treatment but not following vancomycin or metronidazole treatment.<sup>30</sup>
- Gut microbiota has a significant impact on *C. rodentium* disease course.<sup>30</sup>
- Age, host genetic background, diet, and indigenous microbiota influence disease expression.<sup>9</sup>
- Commercial diets effect the baseline colon morphology and presumably the epithelial cell turnover rate – the dietary constituents responsible for this effect are unknown.<sup>8</sup>

## Pathology

- Mice develop colitis following infection.<sup>31</sup>
- Hallmark pathologic lesion: colonic hyperplasia with limited inflammation and epithelial cell hyperproliferation in the descending colon.<sup>26</sup>
- Characterized by crypt elongation, increased mitotic activity, mucosal thickening, variable mucosal inflammation, crypt abscesses, occasional erosions and ulcers, healing, and goblet cell hyperplasia<sup>26,28</sup>
- Grossly thickened, opaque and rigid distal colon, devoid of formed faeces<sup>9,26</sup>
- With increasing severity of disease, the entire colon, and less frequently, the caecum and ileum may be involved.<sup>26</sup>
- Caecum is often empty and contracted.<sup>26</sup>

- Hyperplasia is detectable as early as 4 days after infection, increases in severity through weeks 2 and 3, and resolves 5-8 weeks after infection.<sup>26</sup>
- Peak hyperplastic response occurs within 2-3 weeks with most prominent clinical signs.<sup>7</sup>
- Hyperplasia regresses with excessive goblet cell differentiation and development of cryptal cysts filled with mucin, mucosa returns to normal by 2 months.<sup>8</sup>
- Mucosal hyperplasia is more severe in outbred NIH Swiss mice as compared with C3H/HeJ, C57BL/6J and DBA/2J mice.<sup>8</sup>
- C3H/HeJ mice are more susceptible to disease than DBA/2J or C57BL/6J mice and develop a moderate degree of mucosal hyperplasia.<sup>8</sup>
- Animals of some inbred strains and transgenic lines develop lesions as severe as those seen in suckling mice: neutrophil infiltration of mucosa and submucosa, mucosal erosions, and necrosis.<sup>26</sup>
- Necrosis of the colonic mucosa and severe colitis most notably in suckling mice<sup>29</sup>
- Young mice and certain mouse strains (C3H substrains, AKR, FVB) tend to develop more severe disease with varying mortality due to secondary inflammatory and ulcerative lesions in the hyperplastic mucosa.<sup>7</sup>
- C57BL/6 mice depleted of CD4+ T cells are highly susceptible to infection and develop severe colitis.<sup>21</sup>
- Mice lacking CD4+ T cells or B cells show hypersusceptibility to mucosal inflammation, colonic pathology, and systemic dissemination of *C. rodentium*.<sup>20,32</sup>
- LPS-hyporesponsive C3H/HeJ mice experience more rapid and extensive bacterial colonization than SCID mice – high bacterial load is associated with accelerated crypt hyperplasia, mucosal ulceration, and bleeding, together with very high mortality rates<sup>21</sup>
- Infection of TLR2-deficient mice leads to severe colonic pathology, rapid weight loss and accelerated mortality.<sup>33</sup>
- Caspase 1-deficient mice and NLRP3-deficient mice have increased bacterial loads, aberrant inflammatory and chemokine responses, severe immunopathology and rapid weight loss.<sup>10</sup>
- Mucosal hyperplasia is dependent on the host immune response.<sup>34</sup>
- Hyperplastic responses do not occur in interferon (IFN)- $\gamma$  receptor-deficient mice.<sup>34</sup>
- Depletion of IFN- $\gamma$  prevents crypt hyperplasia.<sup>35</sup>
- CD4-/- or TCR- $\beta$ -/- mice develop polymicrobial sepsis and end-organ damage (abscesses) and succumb during acute infection.<sup>36</sup>
- Innate immunity can mediate acute responses to infection, but T and/or B lymphocytes mediate most of the tissue pathology and inflammation in the later stages of infection.<sup>20</sup>
- Mice devoid of B and T cells, CD4 T cells, or B cells fail to clear infection and succumb from sepsis, with bacteraemia arising from *C. rodentium* and other gut bacteria.<sup>7</sup>
- Bacteraemia and extra-intestinal infection are not hallmarks of infection, though recovery of bacteria from blood, liver and spleen has been reported.<sup>9</sup>
- B cell-deficient (MuMT<sup>-/-</sup>) or B and T cell-deficient (recombinase-activating gene 2<sup>-/-</sup>) mice develop severe pathology in the colon and internal organs and deteriorate rapidly during acute infection.<sup>36</sup>

- Inflammatory and crypt hyperplastic responses in RAG1<sup>-/-</sup> mice are transient and infection is often fatal.<sup>20</sup>
- RAG1<sup>-/-</sup> mice respond to infection primarily with a granulocytic infiltration of the colonic mucosa.<sup>20</sup>

### **Morbidity and mortality**

- Low morbidity and mortality in most adult mice, while mortality is observed in weaning-age mice.<sup>8,26</sup>
- Mortality is significantly higher in C3H/HeJ than in DBA/2J, Swiss, or C57BL/6 mice.<sup>8</sup>
- Increased level of mortality accompanied by a high incidence of rectal prolapse in outbred Swiss-Webster mouse<sup>37</sup>
- High mortality in T-cell receptor  $\alpha\beta$  transgenic mice<sup>17</sup>
- Mortality occurs from bacteraemia, with multifocal hepatitis and splenitis in mice that are incapable of mounting effective immunity.<sup>7</sup>
- Severity of hyperplasia does not correlate with mortality: C3H/HeJ mice did not develop more severe hyperplasia as compared to outbred Swiss-Webster mice, but C3H/HeJ mice did exhibit 45% mortality while no mortality was observed in Swiss-Webster mice.<sup>9</sup>

### **Zoonotic potential**

- No data

### **Interference with research**

#### **Oncology**

- *C. rodentium*-induced hyperplasia can alter chemical carcinogenesis in the large bowel.<sup>8,38</sup>
- Hyperplastic state of the colon serves as a promoter for colon tumorigenesis.<sup>8</sup>
- Transient hyperplastic state increases susceptibility to the carcinogenic effect of 1,2-dimethylhydrazine (DMH) in NIH Swiss mice: epithelial cell hyperproliferation promotes the development of colonic adenomas after administration of DMH.<sup>39</sup>
- DMH administration concomitant with hyperplasia reduces the latency period for early neoplastic lesions, however hyperplasia has no effect on already established tumors.<sup>39</sup>
- Hyperplastic lesions may be confused with neoplasia because associated cytokinetic alterations share several common features with those observed in neoplasia.<sup>40,41</sup>
- Apc<sup>Min/+</sup> mice infected with *C. rodentium* at 1 month of age have a 4-fold increase in the number of colonic adenomas in the distal colon at 6 months of age.<sup>42</sup>

#### **Teratology**

- No data

#### **Infectiology**

- No data

## **Immunology**

- Colonic hyperproliferation is associated with cytokinetic alterations.<sup>40</sup>
- Expression of many proinflammatory cytokines is highly induced in response to *C. rodentium* infection.<sup>43</sup>
- *C. rodentium* infection causes a significant increase in the number of natural T<sub>reg</sub> cells in the colon during active disease.<sup>43</sup>
- Infection generates a predominately lymphocytic infiltrate, characterized by CD4+ T cells situated near the proliferative epithelial crypts.<sup>44</sup>
- Increase of IL-10 secretion and inhibition of IL-2 and IL-4 secretion by mitogen-stimulated murine spleen cells<sup>45</sup>
- Variable effect on IFN- $\gamma$  secretion, whereas the effect of enteropathogenic *Escherichia coli* lysates is inhibitory<sup>45</sup>
- Suppression of lymphocyte activation in vitro<sup>46</sup>
- Highly polarized Th 1 immune response in the gut, characterized by increased levels of IL-12, IFN- $\gamma$  and TNF- $\alpha$  mRNA<sup>44</sup>
- *C. rodentium* triggers a potent Th17 cell response 1 week after oral challenge.<sup>10</sup>
- Infected IL-12p40<sup>-/-</sup> and IFN- $\gamma$ <sup>-/-</sup> mice mount anti-*Citrobacter* serum and gut associated IgA responses and strongly express inducible NO synthase (iNOS) in mucosal tissue, despite diminished serum nitrite/nitrate levels.<sup>19</sup>
- Up-regulated expression of the mouse  $\beta$ -defensins (mBD)-1 and mBD-3 in colonic tissue in C57BL/6 mice; in contrast, only up-regulated expression of mBD-3 in IL-12- and IFN- $\gamma$ -deficient mice<sup>19</sup>
- MYD88 signalling controls *C. rodentium* infection: MYD88 deficiency compromises the ability of the host to restrict bacterial replication<sup>10</sup>
- Increased IL-17 and segmented filamentous bacteria (SFB) levels correlate with increased resistance to *C. rodentium*.<sup>47</sup>

## **Interactions with other infectious agents**

- Probiotics are able to prevent *C. rodentium*-induced mortality in neonatal mice and are able to reduce *C. rodentium* colonization.<sup>10</sup>
- *Bacillus subtilis* slightly reduces colonic hyperplasia due to *C. rodentium* infection when administered to suckling Swiss NIH mice.<sup>48</sup>
- *Lactobacillus acidophilus* NCFM reduces colonic hyperplasia and inflammatory infiltrates due to *C. rodentium* infection in suckling BALB/cByJ mice.<sup>49</sup>
- Administration of a probiotic cocktail containing *Lactobacillus acidophilus* and *Lactobacillus rhamnosus* in C57BL/6J mice enhances bacterial killing, prevention of bacterial attachment, or enhancement of the Th1/T-regulatory response.<sup>50</sup>
- Coinfection with the intestinal helminth *Heligmosomoides polygyrus* can promote *C. rodentium*-associated disease and colitis in BALB/c mice and displays a marked morbidity and mortality.<sup>51</sup>
- Persistent *Helicobacter hepaticus* infection causes chronic *C. rodentium*-induced colitis in C57BL/6J mice, resulting in delayed recovery from weight loss and tissue damage.<sup>43</sup>
- *C. rodentium* infection modulates the outcome of fungal infection in the lung with *Cryptococcus neoformans* in C57BL/6 mice.<sup>52</sup>

- *C. rodentium* infection causes a pronounced dysbiosis that is characterized by an overgrowth of *C. rodentium* and a reduction in the abundance and overall diversity of the resident microbiota.<sup>53</sup>
- Transfer of the microbiota from resistant mice to mice that are susceptible to infection results in the transfer of host resistance to *C. rodentium* infection.<sup>10</sup>

### **Toxicology**

- No data

### **Physiology**

- *C. rodentium* infection causes alterations of phase I and phase II metabolic enzymes in liver and kidney, as well as increases in hepatic cytokine transcripts; distinct liver pathology correlated with serum elevation of liver transaminases, systemic and hepatic cytokine, and chemokine changes in C57BL/6 mice.<sup>54</sup>

### **Cell biology**

- Increase in total cellular  $\beta$ -catenin accompanied by an increase in nuclear  $\beta$ -catenin concentrations; elevated levels preceded crypt elongation<sup>55</sup>
- Increase in cellular concentrations of cyclin D1 and c-Myc (proteins maintaining proliferation status)<sup>55</sup>
- Increased transcription of EGR-1 with subsequent activation of the MEK/extracellular signal-regulated kinases<sup>56</sup>
- Increase in production of keratinocyte growth factor, which induces cell proliferation<sup>44,57</sup>

### **Assisted reproductive technology**

- No data

### **Special considerations**

- The *C. rodentium* mouse model is the gold standard for studying virulence of the closely related human attaching and effacing enteropathogenic *Escheria coli* (EPEC) and enterohemorrhagic *Escheria coli* (EHEC).<sup>10,23,58</sup>
- *C. rodentium* infection is used to model several human intestinal disorders like Crohn's disease, ulcerative colitis, and colon tumorigenesis.<sup>10</sup>
- *C. rodentium* is a tool to investigate adaptive mucosal immune responses to bacterial infection.<sup>10,23</sup>
- *C. rodentium* infection is a reproducible, robust, and physiologically relevant model of inflammation.<sup>54</sup>



## References

1. Coleman GL, Bhatt PN, Jonas AM. 1972. Hyperplastic colitis in mice. 23rd Annual Meeting, AALAS, St. Louis.
2. Barthold SW, Coleman GL, Bhatt PN, Osbaldiston GW, Jonas AM. 1976 The etiology of transmissible murine colonic hyperplasia. *Lab Anim Sci* 26:889-894.
3. Brenner DJ, Grimont PA, Steigerwalt AG, Fanning GR, Ageron E, Riddle CF. 1993. Classification of *Citrobacter* by DNA hybridization: designation of *Citrobacter farmeri* sp. nov., *Citrobacter youngae* sp. nov., *Citrobacter braakii* sp. nov., *Citrobacter werkmanii* sp. nov., *Citrobacter sedlakii* sp. nov., and three unnamed *Citrobacter* genomospecies. *Int J Syst Bacteriol* 43(4):645-658.
4. Schauer DB, Falkow S. 1993. Attaching and effacing locus of a *Citrobacter freundii* biotype that causes transmissible murine colonic hyperplasia. *Infect Immun* 61(6):2486-2492.
5. Schauer DB, Zabel BA, Pedraza IF, O'Hara CM, Steigerwalt AG, Brenner DJ. 1995. Genetic and biochemical characterization of *Citrobacter rodentium* sp. nov. *J Clin Microbiol* 33(8):2064-2068.
6. Barthold SW, Osbaldiston GW, Jonas AM. 1977. Dietary, bacterial and host genetic interactions in the pathogenesis of transmissible murine colonic hyperplasia. *Lab Anim Sci* 27(6):938-945.
7. Luperchio SA, Schauer DB. 2001. Molecular pathogenesis of *Citrobacter rodentium* and transmissible murine colonic hyperplasia. *Microb Infect* 3(4):333-340.
8. Pritchett-Corning KR, Cosentino J, Clifford CB. 2009. Contemporary prevalence of infectious agents in laboratory mice and rats. *Lab Anim* 43(2):165-173.
9. Barthold SM, Griffey SM, Percy DH (eds). 2016. Pathology of laboratory rodents and rabbits, 4th ed. Ames: Wiley Blackwell, pp.50-52.
10. Collins JW, Keeney KM, Crepin VF, Rathinam VA, Fitzgerald KA, Finlay BB, Frankel G. 2014. *Citrobacter rodentium*: infection, inflammation and the microbiota. *Nat Rev Microbiol* 12(9):612-623.
11. Deng W, Li Y, Vallance BA, Finlay BB. 2001. Locus of enterocyte effacement from *Citrobacter rodentium*: sequence analysis and evidence for horizontal transfer among attaching and effacing pathogens. *Infect Immun* 69(10):6323-6335.
12. Frankel G, Phillips AD, Novakova M, Field H, Candy DC, Schauer DB, Douce G, Dougan G. 1996. Intimin from enteropathogenic *Escherichia coli* restores murine virulence to a *Citrobacter rodentium* eaeA mutant: induction of an immunoglobulin A response to intimin and EspB. *Infect Immun* 64(12):5315-5325.
13. Barthold SW. 1980. The microbiology of transmissible murine colonic hyperplasia. *Lab Anim Sci* 30:167-173.
14. Itoh K, Matsui T, Tsuji K, Mitsuoka T, Ueda K. 1988. Genetic control in the susceptibility of germfree inbred mice to infection by *Escherichia coli* O115a,c:K(B). *Infect Immun* 56(4):930-935.
15. Silverman J, Chavannes JM, Rigotty J, Ornaf M. 1979. A natural outbreak of transmissible murine colonic hyperplasia in A/J mice. *Lab Anim Sci* 29(2):209-213.
16. Bieniek H, Tober-Meyer B. 1976. Etiology of colitis and of rectal prolapse in the mouse. *Z Versuchstierkd* 18(5-6):337-348.
17. Maggio-Price L, Nicholson KL, Kline KM, Birkebak T, Suzuki I, Wilson DL, Schauer D, Fink PJ. 1998. Diminished reproduction, failure to thrive, and altered immunologic function in a colony of T-cell receptor transgenic mice: possible role of *Citrobacter rodentium*. *Lab Anim Sci* 48(2):145-155.

18. Goncalves NS, Ghaem-Maghani M, Monteleone G, Frankel G, Dougan G, Lewis DJ, Simmons CP, MacDonald TT. 2001. Critical role for tumor necrosis factor alpha in controlling the number of luminal pathogenic bacteria and immunopathology in infectious colitis. *Infect Immun* 69(11):6651-6659.
19. Simmons CP, Goncalves NS, Ghaem-Maghani M, Bajaj-Elliott M, Clare S, Neves B, Frankel G, Dougan G, MacDonald TT. 2002. Impaired resistance and enhanced pathology during infection with a noninvasive, attaching-effacing enteric bacterial pathogen, *Citrobacter rodentium*, in mice lacking IL-12 or IFN-gamma. *J Immunol* 168(4):1804-1812.
20. Vallance BA, Deng W, Knodler LA, Finlay BB. 2002. Mice lacking T and B lymphocytes develop transient colitis and crypt hyperplasia yet suffer impaired bacterial clearance during *Citrobacter rodentium* infection. *Infect Immun* 70(4):2070-2081.
21. Vallance BA, Deng W, Jacobson K, Finlay BB. 2003. Host susceptibility to the attaching and effacing bacterial pathogen *Citrobacter rodentium*. *Infect Immun* 71(6):3443-3453.
22. MacDonald TT, Frankel G, Dougan G, Goncalves NS, Simmons C. 2003. Host defences to *Citrobacter rodentium*. *Int J Med Microbiol* 293(1):87-93.
23. Borenshtein D, McBee ME, Schauer DB. 2008. Utility of the *Citrobacter rodentium* infection model in laboratory mice. *Curr Opin Gastroenterol* 24(1):32-37.
24. Bergstrom KS, Kisson-Singh V, Gibson DL, Ma C, Montero M, Sham HP, Ryz N, Huang T, Velcich A, Finlay BB, Chadee K, Vallance BA. 2010. Muc2 protects against lethal infectious colitis by disassociating pathogenic and commensal bacteria from the colonic mucosa. *PLoS Pathog* 6(5):e1000902.
25. de la Puente-Redondo VA, Gutiérrez-Martín CB, Pérez-Martínez C, del Blanco NG, García-Iglesias MJ, Pérez-García CC, Rodríguez-Ferri EF. 1999. Epidemic infection caused by *Citrobacter rodentium* in a gerbil colony. *Vet Rec* 145(14):400-403.
26. Barthold SW, Coleman GL, Jacoby RO, Livestone EM, Jonas AM. 1978. Transmissible murine colonic hyperplasia. *Vet Path* 15(2):223-236.
27. Wiles S, Clare S, Harker J, Huett A, Young D, Dougan G, Frankel G. 2004. Organ specificity, colonization and clearance dynamics in vivo following oral challenges with the murine pathogen *Citrobacter rodentium*. *Cell Microbiol* 6(10):963-972.
28. National Research Council. 1991. Committee on Infectious Diseases of Mice and Rats. Infectious diseases of mice and rats. Washington, DC: National Academy Press, pp. 139-141.
29. Luperchio SA, Newman JV, Dangler CA, Schrenzel MD, Brenner DJ, Steigerwalt AG, Schauer DB. 2000. *Citrobacter rodentium*, the causative agent of transmissible murine colonic hyperplasia, exhibits clonality: synonymy of *C. rodentium* and mouse pathogenic *Escherichia coli*. *J Clin Microbiol* 38(12):4343-4350.
30. Mullineaux-Sanders C, Collins JW, Ruano-Gallego D, Levy M, Pevsner-Fischer M, Glegola-Madejska IT, Sågfors AM, Wong JLC, Elinav E, Crepin VF, Frankel G. 2017. *Citrobacter rodentium* relies on commensals for colonization of the colonic mucosa. *Cell Rep* 21(12):3381-3389.
31. Mundy R, MacDonald TT, Dougan G, Frankel G, Wiles S. 2005. *Citrobacter rodentium* of mice and man. *Cell Microbiol* 7(12):1697-1706.
32. Simmons CP, Clare S, Ghaem-Maghani M, Uren TK, Rankin J, Huett A, Goldin R, Lewis DJ, MacDonald TT, Strugnell RA, Frankel G, Dougan G. 2003. Central role for B lymphocytes and CD4+ T cells in immunity to infection by the attaching and effacing pathogen *Citrobacter rodentium*. *Infect Immun* 71(9):5077-5086.

33. Gibson DL, Ma C, Rosenberger CM, Bergstrom KS, Valdez Y, Huang JT, Khan MA, Vallance BA. 2008. Toll-like receptor 2 plays a critical role in maintaining mucosal integrity during *Citrobacter rodentium*-induced colitis. *Cell Microbiol* 10(2):388-403.
34. Higgins LM, Frankel G, Connerton I, Gonçalves NS, Dougan G, MacDonald TT. 1999. Role of bacterial intimin in colonic hyperplasia and inflammation. *Science* 285(5427):588-591.
35. Artis D, Potten CS, Else KJ, Finkelman FD, Grencis RK. 1999. *Trichuris muris*: host intestinal epithelial cell hyperproliferation during chronic infection is regulated by interferon-gamma. *Exp Parasitol* 92(2):144-153.
36. Bry L, Brenner MB. 2004. Critical role of T cell-dependent serum antibody, but not the gut-associated lymphoid tissue, for surviving acute mucosal infection with *Citrobacter rodentium*, an attaching and effacing pathogen. *J Immunol* 172(1):433-441.
37. Ediger RD, Kovatch RM, Rabstein MM. 1974. Colitis in mice with a high incidence of rectal prolapse. *Lab Anim Sci* 24(3):488-494.
38. Barthold SW, Beck D. 1980. Modification of early dimethylhydrazine carcinogenesis by colonic mucosal hyperplasia. *Cancer Res* 40(12):4451-4455.
39. Barthold SW, Jonas AM. 1977. Morphogenesis of early 1,2-dimethylhydrazine-induced lesions and latent reduction of colonic carcinogenesis in mice by variant of *Citrobacter freundii*. *Cancer Res* 37(12): 4352-4360.
40. Barthold SW. 1979. Autoradiographic cytokinetics of colonic mucosal hyperplasia in mice. *Cancer Res* 39(1):24-29.
41. Pulliger BD, Iversen S. 1960. Mammary tumor incidence in relation to age and number of litters in C3Hf and RIII f mice. *Br J Cancer* 14:267-278.
42. Newman JV, Kosaka T, Sheppard BJ, Fox JG, Schauer DB. 2001. Bacterial infection promotes colon tumorigenesis in *Apc*(Min/+) mice. *J Infect Dis* 184(2):227-230.
43. McBee ME, Zheng PZ, Rogers AB, Fox JG, Schauer DB. 2008. Modulation of acute diarrheal illness by persistent bacterial infection. *Infect Immun* 76(11):4851-4858.
44. Higgins LM, Frankel G, Douce G, Dougan G, MacDonald TT. 1999. *Citrobacter rodentium* infection in mice elicits a mucosal Th1 cytokine response and lesions similar to those in murine inflammatory bowel disease. *Infect Immun* 67(6):3031-3039.
45. Malstrom C, James S. 1998. Inhibition of murine splenic and mucosal lymphocyte function by enteric bacterial products. *Infect Immun* 66(7): 3120-3127.
46. Klapproth JMA, Scaletsky ICA, McNamara BP, Lai LC, Malstrom C, James SP, Donnenberg MS. 2000. A large toxin from pathogenic *Escherichia coli* strains that inhibits lymphocyte activation. *Infect Immun* 68(4):2148-2155.
47. Ivanov II, Atarashi K, Manel N, Brodie EL, Shima T, Karaoz U, Wei D, Goldfarb KC, Santee CA, Lynch SV, Tanoue T, Imaoka A, Itoh K, Takeda K, Umesaki Y, Honda K, Littman DR. 2009. Induction of intestinal Th17 cells by segmented filamentous bacteria. *Cell* 139(3):485-498.
48. D'Arienzo R, Maurano F, Mazzarella G, Luongo D, Stefanile R, Ricca E, Rossi M. 2006. *Bacillus subtilis* spores reduce susceptibility to *Citrobacter rodentium*-mediated enteropathy in a mouse model. *Res Microbiol* 157(9):891-897.
49. Chen CC, Louie S, Shi HN, Walker WA. 2005. Preinoculation with the probiotic *Lactobacillus acidophilus* early in life effectively inhibits murine *Citrobacter rodentium* colitis. *Pediatr Res* 58(6):1185-1191.

50. Johnson-Henry KC, Nadjafi M, Avitzur Y, Mitchell DJ, Ngan BY, Galindo-Mata E, Jones NL, Sherman PM. 2005. Amelioration of the effects of *Citrobacter rodentium* infection in mice by pretreatment with probiotics. *J Infect Dis* 191(12):2106-2117.
51. Chen CC, Louie S, McCormick B, McCormick B, Walker WA, Shi HN. 2005. Concurrent infection with an intestinal helminth parasite impairs host resistance to enteric *Citrobacter rodentium* and enhances *Citrobacter*-induced colitis in mice. *Infect Immun* 73(9):5468-5481.
52. Williams AE, Edwards L, Hussell T. 2006. Colonic bacterial infection abrogates eosinophilic pulmonary disease. *J Infect Dis* 193(2):223-230.
53. Lupp C, Robertson ML, Wickham ME, Sekirov I, Champion OL, Gaynor EC, Finlay BB. 2007. Host-mediated inflammation disrupts the intestinal microbiota and promotes the overgrowth of *Enterobacteriaceae*. *Cell Host Microbe* 2(2):119-129.
54. Raczynski AR, Muthupalani S, Schlieper K, Fox JG, Tannenbaum SR, Schauer DB. 2012. Enteric infection with *Citrobacter rodentium* induces coagulative liver necrosis and hepatic inflammation prior to peak infection and colonic disease. *PLoS One* 7(3):e33099.
55. Sellin JH, Umar S, Xiao J, Morris AP. 2001. Increased beta-catenin expression and nuclear translocation accompany cellular hyperproliferation *in vivo*. *Cancer Res* 61(7):2899-2906.
56. de Grado M, Rosenberger CM, Gauthier A, Vallance BA, Finlay BB. 2001. Enteropathogenic *Escherichia coli* infection induces expression of early growth response factor by activating mitogen-activated protein kinase cascades in epithelial cells. *Infect Immun* 69(10):6217-6224.
57. Bajaj-Elliott M, Poulosom R, Pender SL, Wathen NC, MacDonald TT. 1998. Interactions between stromal cell-derived keratinocyte growth factor and epithelial transforming growth factor in immune-mediated crypt cell hyperplasia. *J Clin Invest* 102(8):1473-1480.
58. Crepin VF, Collins JW, Habibzay M, Frankel G. 2016. *Citrobacter rodentium* mouse model of bacterial infection. *Nat Protoc* 11(10):1851-1876.

### **Disclaimer**

Any use of GV-SOLAS booklets (publications) and statements and the application of the information contained therein are at the express risk of the user. Neither GV-SOLAS nor the authors can accept liability for any accidents or damages of any kind arising from the use of a publication (e.g., resulting from the absence of safety instructions), irrespective of legal grounds. Liability claims against GV-SOLAS and the author for damages of a material or non-material nature caused by the use or non-use of the information or by the use of erroneous and/or incomplete information are in principle excluded. Legal claims and claims for damages are thus excluded. The work, including all content, has been compiled with utmost care. However, GV-SOLAS and the authors assume no responsibility for the currentness, correctness, completeness or quality of the information provided. Printing errors and incorrect information cannot be completely ruled out. GV-SOLAS and the authors accept no liability for the currentness, correctness and completeness of the content of the publications or for printing errors. GV-SOLAS and the authors accept no legal responsibility or liability in any form for incorrect statements and consequences arising therefrom. Responsibility for the content of the internet pages printed in these publications lies solely with the owner of the websites concerned. GV-SOLAS and the authors have no influence on the design and content of third-party websites. GV-SOLAS and the authors therefore distance themselves from all third-party content. Responsibility within the meaning of press legislation lies with the board of GV-SOLAS.