

Expert Information

From the Working Group on Hygiene

Implication of infectious agents on results of animal experiments Corynebacterium bovis

Status February 2021

Authors: GV-SOLAS Working Group on Hygiene

Contents

Background	
Prevalence	3
Host species	3
Properties	3
Susceptibility	3
Organotropism	3
Clinical disease	4
Pathology	4
Morbidity and mortality	4
Zoonotic potential	4
Interference with research	4
Oncology	4
Teratology	4
Infectiology	4
Immunology	4
Interactions with other infectious agents	5
Toxicology	5
Physiology	5
Cell biology	5
Assisted reproductive technology	5
Special considerations	5
References	6

Corynebacterium bovis

Background

- First description and isolation from cattle in 1962¹
- First isolation from (athymic) mice in 1995²

Prevalence

- Corynebacterium (C.) bovis is one of the most prevalent organisms isolated from bovine milk, its pathogenicity regarding bovine mastitis is discussed.^{1,3}
- Monitoring of immunodeficient mice with scaly skin lesions revealed that 2% of microbiological cultures and 9% of PCR assays were positive for Corynebacterium bovis.⁴
- Corynebacterium bovis was also isolated from a chronic testicular abscess and the lung of a laboratory rabbit.⁵
- Up to 4% of cultured cell lines and 13% of solid tumor tissue samples were contaminated with *Corynebacterium bovis* DNA.⁶

Host species

- Mouse^{2,7}, rat⁸, rabbit⁵
- Evidence in mouse colonies is reported for North America², Europe⁹ and Asia¹⁰
- Major host: cattle^{1,3}
- Corynebacterium bovis was identified as the causal agent for hyperkeratotic dermatitis (so-called scaly skin disease) in athymic mice in 1995.²

Properties

- Gram-positive, rod-shaped, non spore-forming, facultative anaerobic bacterium^{11,2,9} with different virulence and pathogenicity of strains^{12,13}
- Transmitted between animals by direct contact⁵, spreads rapidly to naïve nude mice.⁹
- Transmission via fomites, for instance, gloves of animal caretakers or by transplantable tumor lines⁹
- Persistent in the environment, lipophilic, survives well on skin flakes or other organic material, only sensitive to heat and certain chemical disinfectants.⁹
- Very difficult to eradicate^{14,15}

Susceptibility

- Immunocompetent and immunodeficient mice and rats can be infected, both glabrous and hirsute.^{7,8}
- Glabrous mice are prone to develop hyperkeratosis.^{7,9}
- Nude (*Foxn1*^{nu}), hairless (*Hr*^{hr})², SCID (*Prkdc*^{scid}), and NSGS mice develop clinical signs and histological findings.^{9,16}

Organotropism

- Skin^{1,2}
- Mucous membranes of oral cavity and nose²

Clinical disease

- Scaly skin disease of athymic mice has been reported since 1976^{2,13}
- Hyperkeratotic dermatitis (also called scaly skin disease) is most commonly observed in homozygous nude mice.^{2,9}
- Euthymic hairless mice may also develop scaly skin disease.²
- SCID mice may develop alopecic areas, with small white flakes involving the dorsum, flanks, neck and cheeks; acanthosis and hyperkeratosis may be observed histologically.⁷
- NSGS mice develop periocular and facial hyperkeratosis and alopecia.
- Cattle mastitis¹, rabbit abscesses⁵, inflammations in different organs of human¹⁷

Pathology

- Dyskeratotic hair follicles with squamous metaplasia along the hair root¹⁸
- Marked acanthosis, moderate orthokeratotic hyperkeratosis, dermal mononuclear cell infiltrate²

Morbidity and mortality

- Usually inapparent infection for immunocompetent and hirsute animals¹⁹
- High morbidity (80%) but low mortality (1%) in affected adult hairless mice^{7,20}
- Suckling mice have high mortality up to 100%.¹⁸
- The lesions tend to resolve spontaneously and disappear within 7-10 days.^{2,8}

Zoonotic potential

Unclear; some cases of human diseases have been reported. 14,17,21,22

Interference with research

Oncology

- Tumor growth rate may be depressed. 18,20
- Engraftment of xenografts is retarded, NK cell activity may be affected. 18,20
- In the NSGS mouse model of chronic myelomonocytic leukemia (CMML), *C. bovis* infection results in diminished human CMML engraftment, including less thrombocytopenia, less splenomegaly, fewer CMML infiltrates in histopathologic sections of murine organs, and fewer human CD45+ cells in samples from spleen, bone marrow, and peripheral blood.¹⁶

Teratology

No data

Infectiology

No data

Immunology

No data

Interactions with other infectious agents

• Corynebacterium bovis infection of the mammary gland in mice has an influence on subsequent infection with Staphylococcus aureus. 12

Toxicology

No data

Physiology

- Increased water consumption (2x) and urine production^{2,20}
- Significant weight loss¹⁹, probably due to anorexia and dehydration or depressed growth rate

Cell biology

No data

Assisted reproductive technology

No data

Special considerations

No data

Updated by Karin Jacobi, Berlin, March 2019 and Michael Mähler, Hannover, February 2021

References

- 1. Cobb RW, Walley JK. 1962. *Corynebacterium bovis* as a probable cause of bovine mastitis. Vet Rec 74:101-102.
- 2. Clifford CB, Walton BJ, Reed TH, Coyle MB, White WJ, Amyx HL. 1995. Hyperkeratosis in athymic nude mice caused by a coryneform bacterium: microbiology, transmission, clinical signs and pathology. Lab Anim Sci 45(2):131-139.
- 3. Brooks BW, Barnum DA, Meek AH. 1983. An observational study of *Corynebacterium bovis* in selected Ontario Dairy Herds. Can J Comp Med 47(1):73-78.
- 4. Pritchett-Corning KR, Cosentino J, Clifford CB. 2009. Contemporary prevalence of infectious agents in laboratory mice and rats. Lab Anim 43(2):165-173.
- 5. Arseculeratne SN, Navaratnam C. 1975. *Corynebacterium bovis* as a pathogen in rabbits. Res Vet Sci 18(2):216-217.
- Manuel CA, Bagby SM, Reisinger JA, Pugazhenthi U, Pitts TM, Keysar SB, Arcaroli JJ, Leszczynski JK. 2017. Procedure for horizontal transfer of patient-derived xenograft tumors to eliminate Corynebacterium bovis. J AM Assoc Lab Anim Sci 56(2):166-172.
- Scanziani E, Gobbi A, Crippa L, Giusti AM, Pesenti E, Cavalletti E, Luini M. 1998. Hyperkeratosisassociated coryneform infection in severe combined immunodeficient mice. Lab Anim 32(3):330-336.
- 8. Burr HN, Lipman NS, White JR, Zheng J, Wolf FR. 2011. Strategies to prevent, treat, and provoke Corynebacterium-associated hyperkeratosis in athymic nude mice. J Am Assoc Lab Anim Sci 50(3):378-388.
- 9. Scanziani E, Gobbi A, Crippa L, Giusti AM, Giavazzi R, Cavalletti E, Luini M. 1997. Outbreak of hyperkeratotic dermatitis of athymic nude mice in northern Italy. Lab Anim 31(3):206-211.
- 10. Kim TH, Kim DS, Han JH, Chang SN, Kim KS, Seok SH, Kim DJ, Park JH, Park JH. 2014. Detection of *Corynebacterium bovis* infection in athymic nude mice from a research animal facility in Korea. J Vet Sci 15(4):583-586.
- 11. Anderson JC, Honkanen-Buzalski T, Bramley AJ. 1985. The pathogenesis of a high-virulence strain of *Corynebacterium bovis* in the mammary gland of the mouse. J Comp Pathol 95(2):227-234.
- 12. Honkanen-Buzalski T, Anderson JC, Bramley AJ. 1985. The virulence of strains of *Corynebacterium bovis* in the mammary gland of the mouse and the effect of corynebacterial mastitis on subsequent infection with *Staphylococcus aureus*. Br Vet J 141(5):519-528.
- 13. Dole VS, Henderson KS, Fister RD, Pietrowski MT, Maldonado G, Clifford CB. 2013. Pathogenicity and genetic variation of 3 strains of *Corynebacterium bovis* in immunodeficient mice. J Am Assoc Lab Anim Sci 52(4):458-466.
- 14. Burr HN, Wolf FR, Lipman NS. 2012 *Corynebacterium bovis*: epizootiologic features and environmental contamination in an enzootically infected rodent room. Lab Anim Sci 51(2):189-198.
- 15. Manuel CA, Pugazhenti U, Leszczynski JK. 2016. Surveillance of a ventilated rack system for *Corynebacterim bovis* by sampling exhaust-air manifolds. J Am Assoc Lab Anim Sci 55(1):58-65.
- 16. Vedder AR, Miedel EL, Ragland NH, Balasis ME, Letson CT, Engelman RW, Padron E. 2019. Effects of *Corynebacterium bovis* on engraftment of patient-derived chronic myelomonocytic leukemia cells in NSGS mice. Comp Med 64(4): 276-282.
- 17. Vale JA, Scott GW. 1977. *Corynebacterium bovis* as a cause of human disease. Lancet 2(8040):682-684,
- 18. Field G. 2006. An update on scaly skin disease. ACLAM Newsletter 37:5-8.

- 19. Gobbi A, Crippa L, Scanziani E. 1999. *Corynebacterium bovis* infection in immunocompetent hirsute mice. Lab Anim Sci 49(2):209-211.
- 20. Field K, Greenstein G, Smith M, Herrmann S, Gizzi J. 1995. Hyperkeratosis-associated coryneform in athymic nude mice. Lab Anim Sci 45:469 (abstract).
- 21. Dalal A, Urban C, Ahluwalia M, Rubin D. 2008. *Corynebacterium bovis* line related septicemia: a case report and review of the literature. Scand J Infect Diseases 2008; 40(6-7):575-577.
- 22. Sbaai M, Lahlou YB, Ghazouniani M, Houba A, Frikh M, Elouenass M. 2011. Septicémie á *Corynebacterium bovis*: á propos dún cas avec revue de littérature. Ann Biol Clin 69(6):732-734

Disclaimer

Any use of GV-SOLAS booklets (publications) and statements and the application of the information contained therein are at the express risk of the user. Neither GV-SOLAS nor the authors can accept liability for any accidents or damages of any kind arising from the use of a publication (e.g. resulting from the absence of safety instructions), irrespective of legal grounds. Liability claims against GV-SOLAS and the author for damages of a material or non-material nature caused by the use or non-use of the information or by the use of erroneous and/or incomplete information are in principle excluded. Legal claims and claims for damages are thus excluded. The work, including all content, has been compiled with utmost care. However, GV-SOLAS and the authors assume no responsibility for the currentness, correctness, completeness or quality of the information provided. Printing errors and incorrect information cannot be completely ruled out. GV-SOLAS and the authors accept no liability for the currentness, correctness and completeness of the content of the publications or for printing errors. GV-SOLAS and the authors accept no legal responsibility or liability in any form for incorrect statements and consequences arising therefrom. Responsibility for the content of the internet pages printed in these publications lies solely with the owner of the websites concerned. GV-SOLAS and the authors have no influence on the design and content of third-party websites. GV-SOLAS and the authors therefore distance themselves from all third-party content. Responsibility within the meaning of press legislation lies with the board of GV-SOLAS.