

Expert Information

from the Working Group on Hygiene

Implication of infectious agents on results of animal experiments

Toxoplasma gondii (description for intermediate hosts)

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Toxoplasma gondii

Background

 Toxoplasma (T.) gondii was initially described more than 100 years ago in Tunis in the tissues of the gundi (Ctenodoactylus gundi), a North African rodent, and in Brazil in the tissues of a rabbit. T. gondii is a ubiquitous, Apicomplexan parasite of warmblooded animals that can cause several clinical symptomes including encephalitis, chorioretinitis, congenital infection and neonatal mortality. Fifteen years after the description of T. gondii a fatal case of toxoplasmosis in a child was reported. In 1939 T. gondii was for the first time conclusively described as a cause of human disease. It was not until the 1960s and 1970s that the parasite was identified as a coccidian. Cats were identified as the definitive host by several groups working independently.¹

Prevalence

- The prevalence of natural infection in laboratory mouse facilities is negligible, because laboratory mice no longer have access to sporulated cysts shed by infected cats, which were historically the major source for cross-infection.²
- In Uganda, Africa 47% of 85 tested free-range chicken were *Toxoplasma* antibody positive.³
- In Vietnam, *Toxoplasma* antibodies were found in pigs in 75 of 325 (23%) finishers, 63 of 207 (32.3%) sows, and 22 of 55 (40%) boars.⁴
- 46% of 84 free-range chicken in a study conducted in Brazil were *Toxoplasma* antibody positive.⁵
- Antibodies to *T. gondii* were found in 52 of 101 (51.5%) dogs tested in Durango City, Mexico.⁶
- *Toxoplasma* was also found in wildlife animals like the Hawaiian monk seal⁷ or the artic fox (*Vulpes lagopus*).⁸

Host species

- Members of the cat family (Felidae; definitive host).9
- All laboratory and domestic animals, birds and humans are intermediate hosts.¹⁰
- Different host species susceptibility.^{11,12}

Properties

- *T. gondii* is an obligate intracellular parasite capable of infecting almost all warmblooded animals and humans.⁹ The three infectious forms of the parasite are: (1) the rapidly dividing tachyzoite, responsible for systemic invasion during primary infection, (2) the slowly growing bradyzoite associated with chronic infection, and (3) the sporozoite, sexually produced in oocysts.¹¹
- Despite having a broad host range, *Felidae* (mainly house cats) are the only hosts in which sexual development is known to take place. Development of gametocytes within intestinal epithelial cells culminates in fertilization and shedding of infectious oocysts with faeces. These spore-like particles can contaminate water and food and lead to infections of a wide range of animal species. In the intermediate host, the

ability to undergo asexual replication, in the form of fast-growing tachyzoites that replicate within nucleated host cells, allows the parasite to rapidly increase in number and disseminate throughout the body. Following a vigorous immune response, the parasite differentiates into semi-dormant tissue cysts that harbour slow growing bradyzoites. Predation and ingestion of tissue cysts by cats completes the cycle.¹⁰

- Persistence of *T. gondii* may lead to physiological and behavioural consequences in infected rodents, resulting in reduced fear of Felidae, their natural predators^{9,13-18}, as well as decreased¹⁹ or higher activity level.²⁰
- After host cell invasion, parasites replicate quickly (known as tachyzoites) by endogeny that leads subsequently to the lysis of the infected cell and spread to neighbouring cells. The parasite is able to cross the intestinal epithelial barrier, disseminate throughout the body, and localize in the muscle, placenta, brain, or eye. Transmigration across polarized epithelial cells is highly associated with the parasite genotype and virulence. Toxoplasma is also able to cross the blood-brain barrier. Ultimately, immune pressure on the protozoan results in the formation of cysts, containing the slow replicating form (bradyzoite) in neuronal tissues and skeletal muscles to establish latent infection.²¹⁻²³

Susceptibility

- Although wild rodents, including house mice, are relatively resistant, laboratory mice are highly susceptible to infection.²⁴
- Differences in genes within the H-2 and H-13 region correlate with resistance or susceptibility to development of *Toxoplasma* encephalitis in mice.²⁵⁻²⁸
- Age, gender, and pregnancy influence susceptibility to *T. gondii* infection in mice. 2-3 months old hybrid mice are more resistant than 22-24 months old hybrid mice, non-pregnant BALB/c females are more resistant than pregnant BALB/c females, male SCID mice are more resistant than female SCID mice.²⁹⁻³¹

Organotropism

- In definite hosts (*Felidae*) shizogonic and gametogonic stages develop usually in the ileum, although the entire length of the small intestine can be affected.³²
- In intermediate hosts, tissue cysts are predominantly located in the brain and other neural tissues, but they also regularly occur in skeletal muscles, heart, tongue and, at least in some host species, in visceral organs such as lung, liver or kidney.³³

Clinical disease

- Usually inapparent (e.g. in rats, cattle, horses, pigs, goats or sheep).¹²
- Severe symptoms and/or febrile disease occur in some species (e.g. marsupials, monkeys, mice).¹²
- Toxoplasmosis can cause necrosis and granulomatous inflammation in the intestine, mesenteric lymph nodes, eyes, heart, adrenals, spleen, brain, lung, liver, placenta, and muscles.²

Pathology

• Central nervous system: parasite tissue cysts are found in all brain areas of mice, prior studies reporting high numbers located in the amygdala and frontal cortex.³⁴⁻³⁷

- Lesions in immunocompromised mice may lack inflammatory infiltrates and solely consist of small necrotic foci and scattered cysts.^{38,39}
- Human cells are destroyed by active multiplication of *T. gondii*. Infection may result in necrotic foci. Congenital infection often involves retina and brain; focal chorioretinitis may result in impaired vision. Brain involvement in immunosuppressed human patients may lead to large necrotic abscesses.⁴⁰

Morbidity and mortality

- Largely depending on the route of infection, parasite strain and dose, and the immunologic state of the host. Mortality in BALB/c and DBA/2 strains was 100% on day 12 and 13 of infection. A hundred percent of mice of the B10.D2 strain died by day 19. Mortality in C57B1/6J and C3H/Bi mice was intermediate, 87 and 80%, respectively. Mice of the DBA/1 and white SW/SIM strains were most resistant, with a mortality of 67% at day 30 after infection.41
- One to 10 oocysts of the M-7741 and the Aldrin Toxoplasma strains killed orally inoculated mice, whereas up to 1,000 oocysts of the BWM and the S-1 strains caused inapparent infections.42,43
- Thirty weeks after infection, 64% of CBA/Ca mice infected with the ME49 Toxoplasma strain had died. In contrast, none of the mice infected with the DAG strain had died.⁴⁴

Zoonotic potential

- *T. gondii* is found in humans worldwide, under a variety of climates and socioeconomic circumstances. It is estimated that one third of the world's human population is chronically infected with this parasite. T. gondii infections cause significant morbidity and mortality worldwide with a wide spectrum of clinical manifestations both in immunocompromised and immunocompetent hosts. Transmission to humans occurs by ingestion of oocyst-contaminated soil and water, from tissue cysts in undercooked meat, by transplantation, blood transfusion, laboratory accidents, or congenitally.^{11,44,46}
- Acute primary infections in pregnant women can have devastating sequelae and can lead to congenital toxoplasmosis with severe neurologic or ocular manifestations and even death.⁴⁶
- Undercooked meat, especially pork, lamb, and wild game meat, are sources of foodborne transmission for humans. The new trend in the production of free-range organically raised meat could increase the risk of *Toxoplasma gondii* contamination of meat.⁴⁷

Interference with research

Oncology

• The interplay between *T. gondii* infection and tumor development is intriguing and not yet fully understood. Some studies showed that *T. gondii* reversed tumor immune suppression, while some reported the opposite, stating that *T. gondii* infection promoted tumor growth.⁴⁸⁻⁵⁰

Teratology

No data

Infectiology / Interactions with other infectious agents

- Macrophage clearance and killing of *Listeria monocytogenes* and *Salmonella typhimurium* are decreased in mice infected with *T. gondii*.⁵¹
- Infection with murine leukemia virus may lead to reactivation of chronic *T. gondii* infection.^{52,53}
- Mice infected with *T. gondii* are resistant to proliferation of *Cryptococcus neoformans* cells in the brain.⁵⁴

Immunology

- Acute and chronic *T. gondii* infections modulate the immune responses in mice.³⁹
- *T. gondii* is able to induce transient immune downregulation.⁵⁵⁻⁵⁸
- *T. gondii*-infected cells are resistant to multiple inducers of apoptosis.⁵⁹
- Gamma delta T-cells induce expression of heat shock protein 65 in macrophages of mice infected with *T. gondii*, thereby preventing the apoptosis of infected macrophages.⁶⁰
- Intracellular *T.* gondii interferes with the MHC class I and class II antigen presentation pathway of murine macrophages.⁶¹
- CD4+ and CD8+ T-lymphocytes appear to act in concert to prevent reactivation of chronic *T. gondii* infection.⁶²⁻⁶⁴
- NK cell activity and production of IFN gamma are increased during the course of *T. gondii* infection in mice; IFN gamma plays a critical role in preventing cyst rupture and toxoplasmic encephalitis.⁶⁵⁻⁶⁸
- Cytokine levels are elevated in infected humans and in murine models of toxoplasmosis. Several publications describe the immunopathology of *T. gondii* infection in humans.⁶⁹⁻⁷³
- IL-12 is crucial for the generation of both innate resistance mechanisms during the acute phase of infection and T-cell-dependent acquired immunity during the chronic phase.⁷⁴
- Various other cytokines, such as IFN-β, IL-1, IL-4, IL-6, IL-10, TGF-β and TNF-α, are implicated in the pathogenesis of *T. gondii* infection.^{69,75-84}
- Inducible nitric oxide is essential for host control of chronic *T. gondii* infection.⁸⁵
- Innate resistance mechanisms during *T. gondii* infection are reviewed by Alexander et al.⁸⁶; T cell-mediated immunity during *T. gondii* infection is reviewed by Denkers and Gazzinelli.⁸⁷

Toxicology

• T. gondii infection increases toxicity of some drugs in mice (e.g. neostigmine).88

Physiology

• Mice infected with *T. gondii* exhibit ovarian dysfunction with uterine atrophy and thyroidal dysfunction (decline in serum thyroxine levels), probably due to impaired release of hypothalamic releasing.^{35,89,90}

Cell biology

• Cell invasion by *T. gondii* into murine cells involves the concerted action of protein secretion along with actin-based motility.¹⁰

Assisted reproductive technology

No data

Special considerations

• Quarantine and testing of all incoming animals from non-commercial sources is recommended

Updated by Thomas Kolbe, Vienna, November 2024

References

- 1. Weiss LM, Dubey JP. 2009. Toxoplasmosis: A history of clinical observations. Int J Parasitol 39(8):895-901. doi:10.1016/j.ijpara.2009.02.004.
- Whary MT, Baumgarth N, Fox JG, Barthold SW. 2015. Chapter 3 Biology and Diseases of Mice. In: Fox JG, Anderson LC, Otto GM, Pritchett-Corning KR, Whary MT (eds.). Laboratory Animal Medicine, 3rd Edition, Academic Press, p123.
- Lindström I, Sundar N, Lindh J, Kironde F, Kabasa JD, Kwok OCH, Dubey JP, Smith JE. 2008. Isolation and genotyping of *Toxoplasma gondii* from Ugandan chickens reveals frequent multiple infections. Parasitol 135(Pt 1):39-45. doi:10.1017/S0031182007003654.
- 4. Huong LTT, Dubey JP. 2007. Seroprevalence of *Toxoplasma gondii* in pigs from Vietnam. J Parasitol 93(4):951-952. doi:10.1645/GE-1163R.1.
- Dubey JP, Sundar N, Gennari SM, Minervino AHH, da R Farias NA, Ruas JL, dos Santos TRB, Cavalcante GT, Kwok OCH, Su C. 2007a. Biologic and genetic comparison of *Toxoplasma gondii* isolates in free-range chickens from the northern Pará state and the southern state Rio Grande do Sul, Brazil revealed highly diverse and distinct parasite populations. Vet Parasitol 143(2):182-188. doi:10.1016/j.vetpar.2006.08.024.
- Dubey JP, Alvarado-Esquivel C, Liesenfeld O, Herrera-Flores RG, Ramírez-Sánchez BE, González-Herrera A, Martínez-García SA, Bandini LA, Kwok OCH. 2007b. *Neospora caninum* and *Toxoplasma gondii* antibodies in dogs from Durango City, Mexico. J Parasitol 93(5):1033-1035. doi:10.1645/GE-1281R.1.
- 7. Aguirre AA, Keefe TJ, Reif JS, Kashinsky L, Yochem PK, Saliki JT, Stott JL, Goldstein T, Dubey JP, Braun R, Antonelis G. 2007. Infectious disease monitoring of the endangered Hawaiian monk seal. J Wildl Dis 43(2):229-241. doi:10.7589/0090-3558-43.2.229.
- 8. Prestrud KW, Dubey JP, Asbakk K, Fuglei E, Su C. 2008. First isolate of *Toxoplasma gondii* from arctic fox (*Vulpes lagopus*) from Svalbard. Vet Parasitol 151(2-4):110-114. doi:10.1016/j.vetpar.2007.11.011.
- 9. Kaushik M, Knowles SCL, Webster JP. 2014. What makes a feline fatal in *Toxoplasma gondii*'s fatal feline attraction? Infected rats choose wild cats. Integr Comp Biol 54(2):118-128. doi:10.1093/icb/icu060.
- 10. Hunter CA, Sibley LD. 2012. Modulation of innate immunity by *Toxoplasma gondii* virulence effectors. Nat Rev Microbiol 10(11):766-778. doi:10.1038/nrmicro2858.
- 11. Saraf P, Keats Shwab E, Dubey JP, Su C. 2017. On the determination of *Toxoplasma gondii* virulence in mice. Exp Parasitol 174:25-30. doi:10.1016/j.exppara.2017.01.009.
- 12. Innes EA. 1997. Toxoplasmosis: comparative species susceptibility and host immune response. Comp Immunol, Microbiol Infect Dis 20:131-138. doi:10.1016/s0147-9571(96)00038-0.
- 13. Berdoy M, Webster JP, Macdonald DW. 2000. Fatal attraction in rats infected with *Toxoplasma gondii*. Proc Biol Sci 267(1452):1591-1594. doi:10.1098/rspb.2000.1182.
- 14. Webster JP. 2001. Rats, cats, people and parasites: the impact of latent toxoplasmosis on behaviour. Microbes Infect Inst Pasteur 3(12):1037-1045. doi:10.1016/s1286-4579(01)01459-9.
- 15. Lamberton PHL, Donnelly CA, Webster JP. 2008. Specificity of the *Toxoplasma gondii*-altered behaviour to definitive versus non-definitive host predation risk. Parasitol 135(10):1143-1150. doi:10.1017/S0031182008004666.

- 16. Kannan G, Moldovan K, Xiao J-C, Yolken RH, Jones-Brando L, Pletnikov MV. 2010. *Toxoplasma gondii* strain-dependent effects on mouse behaviour. Folia Parasitologica 57(2) 151-155. doi:10.14411/fp.2010.019.
- 17. House PK, Vyas A, Sapolsky R. 2011. Predator cat odors activate sexual arousal pathways in brains of *Toxoplasma gondii* infected rats. PLoS One 6(8):e23277. doi:10.1371/journal.pone.0023277.
- Ingram WM, Goodrich LM, Robey EA, Eisen MB. 2013. Mice infected with low-virulence strains of *Toxoplasma gondii* lose their innate aversion to cat urine, even after extensive parasite clearance. PLoS One 8(9):e75246. doi:10.1371/journal.pone.0075246.
- Gonzalez LE, Rojnik B, Urrea F, Urdaneta H, Petrosino P, Colasante C, Pino S, Hernandez L. 2007. *Toxoplasma gondii* infection lower anxiety as measured in the plus-maze and social interaction tests in rats: A behavioral analysis. Behav Brain Res 177(1):70-79. doi:10.1016/j.bbr.2006.11.012.
- 20. Webster JP, Brunton CF, MacDonald DW. 1994. Effect of *Toxoplasma gondii* upon neophobic behaviour in wild brown rats, *Rattus norvegicus*. Parasitol 109(Pt 1):37-43. doi:10.1017/s003118200007774x.
- 21. Black MW, Boothroyd JC. 2000. Lytic Cycle of *Toxoplasma gondii*. Microbiol Mol Biol Rev 64(3): 607-623. doi:10.1128/mmbr.64.3.607-623.2000.
- 22. Barragan A, Sibley LD. 2002. Transepithelial migration of *Toxoplasma gondii* is linked to parasite motility and virulence. J Exp Med 195(12):1625–1633. doi:10.1084/jem.20020258.
- 23. Munoz M, Liesenfeld O, Heimesaat MM. 2011. Immunology of *Toxoplasma gondii*. Immunol Rev 240:269-285. doi:10.1111/j.1600-065X.2010.00992.x.
- 24. Wang Q, Sibley LD. 2020. Assays for monitoring *Toxoplasma gondii* infectivity in the laboratory mouse. Methods Mol Biol 2071:99-116. doi:10.1007/978-1-4939-9857-9_5.
- 25. Williams DM, Grumet FC, Remington JS. 1978. Genetic control of murine resistance to *Toxoplasma gondii*. Infect Immun 19:416-420. doi:10.1128/iai.19.2.416-420.1978.
- 26. Jones TC, Erb P. 1985. H-2 complex-linked resistance in murine toxoplasmosis. J Infect Dis 151:739-740. doi:10.1093/infdis/151.4.739.
- Suzuki, Y, Joh K, Orellana MA, Konley FK, Remington JS. 1991. A gene(s) within the H-2D region determine development of toxoplasmic encephalitis in mice. Immunology 74:732-739. PMID: 1783431.
- Blackwell JM, Roberts CW, Alexander J. 1993. Influence of genes within the MHC on mortality and brain cyst development in mice infected with *Toxoplasma gondii*: kinetics of immune regulation in BALB H-2 congenic mice. Parasite Immunol 15:317-324. doi:10.1111/j.1365-3024.1993.tb00616.x.
- 29. Johnson LL, Gibson GW, Sayles PC. 1995. Preimmune resistance to *Toxoplasma gondii* in aged and young adult mice. J Parasitol 81:894-899. PMID: 8544060.
- 30. Thouvenin M, Candolfi E, Villard O, Klein JP, Kien T. 1997. Immune response in a murine model of congenital toxoplasmosis: increased susceptibility of pregnant mice and transplacental passage of *Toxoplasma gondii* are type 2-dependent. Parasitologia 39:279-283. PMID: 9802080.
- 31. Walker W, Roberts CW, Ferguson DJ, Jebbari H, Alexander J. 1997. Innate immunity to *Toxoplasma gondii* is influenced by gender and is associated with differences in interleukin-12 and gamma interferon production. Infect Immun 65:1119-1121. doi:10.1128/IAI.65.3.1119-1121.1997.
- 32. Hutchison WM, Dunachie JF, Work K, Siim JC. 1971. The life cycle of the coccidian parasite, *Toxoplasma gondii*, in the domestic cat. Trans R Soc Trop Med Hyg 65(3):380-399. doi:10.1016/0035-9203(71)90018-6.

- 33. Swierzy IJ, Muhammad M, Kroll J, Abelmann A, Tenter AM, Lüder CGK. 2014. *Toxoplasma gondii* within skeletal muscle cells: a critical interplay for food-borne parasite transmission. Int J Parasitol 44(2):91-98. doi:10.1016/j.ijpara.2013.10.001.
- 34. Stahl W, Kaneda Y. 1998. Aetiology of thyroidal dysfunction in murine toxoplasmosis. Parasitology 117:223-227. doi:10.1017/s0031182098003035.
- 35. Kittas S, Kittas C, Paizi-Biza P, Henry L. 1984. A histological and immunohistochemical study of the changes induced in the brains of white mice by infection with *Toxoplasma gondii*. Br J Exp Pathol 65:67-74. PMID: 6365146.
- 36. Ferguson DJ, Graham DI, Hutchinson WM. 1991. Pathological changes in the brains of mice infected with *Toxoplasma gondii*: a histological, imunocytochemical and ultrastructural study. Int J Exp Pathol 72:463-474. PMID: 1883744.
- 37. McConkey GA, Martin HL, Bristow GC, Webster JP. 2013. *Toxoplasma gondii* infection and behaviour location, location, location? J Exp Biol 216(1): 113-119. doi:10.1242/jeb.074153.
- 38. Buxton D. 1980. Experimental infection of athymic mice with *Toxoplasma gondii*. J Med Microbiol 13:307-311. doi:10.1099/00222615-13-2-307.
- 39. Johnson LL. 1992. SCID mouse models of acute and relapsing chronic *Toxoplasma gondii* infections. Infect Immun 60:3719-3724. doi:10.1128/iai.60.9.3719-3724.1992.
- 41. Dubey JP. 1996. *Toxoplasma gondii*. In: Baron S (ed) Medical Microbiology, 4th ed. The University of Texas Medical Branch at Galveston, Chapter 84. PMID: 21413265.
- 41. Araujo FG, Williams DM, Grumet FC, Remington JS. 1976. Strain-dependent differences in murine susceptibility to toxoplasma. Infect Immun 13:1528-1530. doi:10.1128/iai.13.5.1528-1530.1976.
- 42. Dubey JP, Frenkel JK. 1973. Experimental Toxoplasma infection in mice with strains producing oocysts. J Parasitol 59:505-512. PMID: 4576142.
- 43. Fernando MA. 1982. Pathology and pathogenicity, In: Long PL (ed) The Biology of the Coccidia, Baltimore: University Park Press, pp. 287-327.
- 44. Suzuki Y, Conley FK, Remington JS. 1989. Differences in virulence and development of encephalitis during chronic infection vary with the strain of *Toxoplasma gondii*. J Infect Dis 159:790-794. doi:10.1093/infdis/159.4.790.
- 45. Elmore SA, Jones JL, Conrad PA, Patton S, Lindsay DS, Dubey JP. 2010. *Toxoplasma gondii*: epidemiology, feline clinical aspects, and prevention. Trends Parasitol 26(4):190-196. doi:10.1016/j.pt.2010.01.009.
- Layton J, Theiopoulou D-C, Rutenberg D, Elshereye A Zhang Y, Sinnott J, Kim K, Montoya JG, Contopoulos-Ioannidis DG. 2023. Clinical Spectrum, Radiological Findings, and Outcomes of Severe Toxoplasmosis in Immunocompetent Hosts: A Systematic Review. Pathogens 12:543. doi:10.3390/pathogens12040543.
- 47. Jones JL, Dubey JP. 2012. Foodborne Toxoplasmosis. Clin Infect Dis 55(6):845-851. doi:10.1093/cid/cis508.
- 48. Bahwal SA, Chen JJ, E L, Hao T, Chen J, Carruthers VB, Lai J. Zhou X. 2022. Attenuated *Toxoplasma gondii* enhances the antitumor efficacy of anti-PD1 antibody by altering the tumor microenvironment in a pancreatic cancer mouse model. J Cancer Res Clin Oncol 148(10):2743-2757. doi:10.1007/s00432-022-04036-8.
- 49. Lu J, Wei N, Zhu S, Chen X, Gong H, Mi R, Huang Y, Chen Z, Li G. 2022. Exosomes derived from dendritic cells infected with *Toxoplasma gondii* show antitumoral activity in a Mouse Model of Colorectal Cancer. Front Oncol 4:12:899737. doi:10.3389/fonc.2022.899737.

- 50. Song Y, Yuan H, Yang X, Ren Z, Qi S, He H, Zhang X-X, Jiang T, Yuan Z-G. 2024. The opposing effect of acute and chronic *Toxoplasma gondii* infection on tumor development. Parasites and Vectors 17(1):247. doi.org/10.1186/s13071-024-06240-6.
- 51. Wing EJ, Boehmer SM, Christner LK. 1983. *Toxoplasma gondii*: decreased resistance to intracellular bacteria in mice. Exp Parasitol 56:1-8. doi:10.1016/0014-4894(83)90090-5.
- 52. Gazzinelli RT, Hartley JW, Fredrickson TN, Chattopadhyay SK, Sher A, Morse HC 3rd. 1992b. Opportunistic infections and retrovirus-induced immunodeficiency: studies of acute and chronic infections with *Toxoplasma gondii* in mice infected with LP-BM5 Murine Leukemia Viruses. Infect Immun 60, 4394-4401. doi:10.1128/iai.60.10.4394-4401.1992.
- 53. Watanabe H, Suzuki Y, Makino M, Fujiwara M. 1993. *Toxoplasma gondii*: Induction of toxoplasmic encephalitis in mice with chronic infection by inoculation of a murine leukemia virus inducing immunodeficiency. Exp Parasitol 76:39-45. doi:10.1006/expr.1993.1005.
- Aguirre KM, Sayles PC, Gibson GW, Johnson LL. 1996. Resistance to *Cryptococcus neoformansis* associated with an inflammatory response to *Toxoplasma gondii* in the central nervous system of mice. Infect Immun 64:77-82. doi:10.1128/iai.64.1.77-82.1996.
- 55. Nguyen TD, Bigaignon G, Van Broeck J, Vercammen M, Nguyen TN, Delmee M, Turneer M, Wolf F, Coutelier JP. 1998. Acute and chronic phases of *Toxoplasma gondii* infection in mice modulate the host immune responses. Infect Immun 66:2991- 2995. doi:10.1128/IAI.66.6.2991-2995.1998.
- 56. Channon JY, Kasper LH. 1996. *Toxoplasma gondii*-induced immune suppression by human peripheral blood monocytes: role of gamma interferon. Infect Immun 64:1181-1189. doi:10.1128/iai.64.4.1181-1189.1996.
- 57. Denkers EY, Caspar P, Hieny S, Sher A. 1996. *Toxoplasma gondii* infection induces specific nonresponsiveness in lymphocytes bearing the V beta 5 chain of the mouse T cell receptor. J Immunol 156:1089-1094. PMID: 8557983.
- 58. Khan IA, Matsuura T, Kasper LH. 1996. Activation-mediated CD4+ T cell unresponsiveness during acute *Toxoplasma gondii* infection in mice. Int Immunol. 8:887-896. doi:10.1093/intimm/8.6.887.
- 59. Nash PB, Purner MB, Leon RP, Clarke P, Duke RC, Curiel TJ. 1998. *Toxoplasma gondii*-infected cells are resistant to multiple inducers of apoptosis. J Immunol 160:1824-1830. PMID: 9469443.
- Hisaeda H, Sakai T, Ishikawa H, Maekawa Y, Yasutomo K, Good RA, Himeno K. 1997. Heat shock protein 65 induced by gammadelta T cells prevents apoptosis of macrophages and contributes to host defense in mice infected with *Toxoplasma gondii*. J Immunol 159:2375-2381. PMID: 9278328.
- 61. Luder CG, Lang T, Beuerle B, Gross U. 1998. Down-regulation of MHC class II molecules and inability to up-regulate class I molecules in murine macrophages after infection with *Toxoplasma gondii*. Clin Exp Immunol 112:308-316. doi:10.1046/j.1365-2249.1998.00594.x.
- 62. Brown CR, McLeod R. 1990. Class I MHC genes and CD8+ T cells determine cyst number in Toxoplasma gondii infection. J Immunol 145:3438-3441. PMID: 2121825.
- 63. Araujo FG. 1991. Depletion of L3T4+ (CD4+) T lymphocytes prevents development of resistance to *Toxoplasma gondii* in mice. Infect Immun 59:1614-1619. doi:10.1128/iai.59.5.1614-1619.1991.
- 64. Gazzinelli RT, Xu Y, Hieny S, Cheever A, Sher A. 1992c. Simultaneous depletion of CD4+ and CD8+ T lymphocytes is required to reactivate chronic infection with *Toxoplasma gondii*. J Immunol 149:175-180. PMID: 1351500.
- 65. Hauser WE, Sharma SD, Remington JS. 1982. Natural killer cells induced by acute and chronic Toxoplasma infection. Cell Immunol 69:330-346. doi:10.1016/0008-8749(82)90076-4.
- 66. Suzuki Y, Conley FK, Remington JS. 1989b. Importance of endogeneous IFN-g for prevention of toxoplasmic encephalitis in mice. J Immunol 143:2045-2050. PMID: 2506275.

- 67. Sher A, Oswald IP, Hieny S, Gazzinelli RT. 1993. *Toxoplasma gondii* induces a T-independent IFNgamma response in natural killer cells that requires both adherent accessory cells and tumor necrosis factor-alpha. J Immunol 150:3982-3989. PMID: 8473745.
- 68. Hunter CA, Subauste C, Remington JS. 1994a. Production of gamma interferon by natural killer cells from *Toxoplasma gondii* infected SCID mice: regulation by interleukin-10, interleukin-12 and tumor necrosis factor alpha. Infect Immun 62:2818-2824. doi:10.1128/iai.62.7.2818-2824.1994.
- 69. Beaman MH, Wong SY, Remington JS. 1992. Cytokines, Toxoplasma and intracellular parasitism. Immunol Rev 127:97-117. doi:10.1111/j.1600-065x.1992.tb01410.x.
- 70. Gazzinelli RT, Denkers EY, Sher A. 1993. Host resistance to *Toxoplasma gondii*: model for studying the selective induction of cell-mediated immunity by intracellular parasites. Infect Agents Dis 2:139-149. PMID: 7909708.
- 71. Subauste CS, Remington JS. 1993. Immunity to *Toxoplasma gondii*. Curr Opin Immunol 5:532-537. doi:10.1016/0952-7915(93)90034-p.
- 72. Hunter CA, Remington JS. 1994. Immunopathogenesis of toxoplasmic encephalitis. J Infect Dis 170:1057-1067. doi:10.1093/infdis/170.5.1057.
- 73. Hunter CA, Subauste C, Remington JS. 1994b. The role of cytokines in toxoplasmosis. Biotherapy 7:237-247. doi:10.1007/BF01878489.
- 74. Johnson LL, Sayles PC. 1997. Interleukin-12, dendritic cells, and the initiation of host-protective mechanisms against *Toxoplasma gondii*. J Exp Med 186:1799-1802. doi:10.1084/jem.186.11.1799.
- 75. Chang HR, Grau GE, Pechere JC. 1990. Role of TNF and IL-1 in infection with *Toxoplasma gondii*. Immunology 69:33-37. PMID: 2107144.
- 76. Orellana MA, Suzuki Y, Araujo FG, Remington JS. 1991. Role of beta interferon in resistance to *Toxoplasma gondii* infection. Infect Immun 59:3287-3290. doi:10.1128/iai.59.9.3287-3290.1991.
- Gazzinelli RT, Oswald IP, Jamos SL, Sher A. 1992a. IL-10 inhibits parasite killing and nitrogen oxide production by IFN-gamma-activated macrophages. J Immunol 148:1792-1796. PMID: 1541819.
- 78. Hunter CA, Bermudez L, Beernink H, Waegell W, Remington JS. 1995a. Transforming growth factor-b inhibits interleukin-12 induced production of interferon-g by natural killer cells: a role for transforming growth factor-b in the regulation of T cell-independent resistance to *Toxoplasma gondii*. Eur J Immunol 25:994-1000. doi:10.1002/eji.1830250420.
- Hunter CA, Chizzonite R, Remington JS. 1995b. IL-1 beta is required for IL-12 to induce production of IFN-gamma by NK cells. A role for IL-1 beta in the T cell-independent mechanism of resistance against intracellular pathogens. J Immunol 155:4347-4354. PMID: 7594594.
- 80. Roberts CW, Ferguson DJ, Jebbari H, Satoskar A, Bluethmann H, Alexander J. 1996. Different roles for interleukin-4 during the course of *Toxoplasma gondii* infection. Infect Immun 64:897-904. doi:10.1128/iai.64.3.897-904.1996.
- Bessieres MH, Swierczynski B, Cassaing S, Miedouge M, Olle P, Seguela JP, Pipy B. 1997. Role of IFN-gamma, TNF-alpha, IL4 and IL10 in the regulation of experimental *Toxoplasma gondii* infection. J Eukaryot Microbiol 44:87S. doi:10.1111/j.1550-7408.1997.tb05800.x.
- Neyer LE, Grunig G, Fort M, Remington JS, Rennick D, Hunter CA. 1997. Role of interleukin-10 in regulation of T-cell-dependent and T-cell-independent mechanisms of resistance to *Toxoplasma gondii*. Infect Immun 65:1675-1682. doi:10.1128/iai.65.5.1675-1682.1997.

- Deckert-Schluter M, Bluethmann H, Rang A, Hof H, Schlüter D. 1998. Crucial role of TNF receptor type 1 (p55), but not of TNF receptor type 2 (p75), in murine toxoplasmosis. J Immunol 160:3427-3436. PMID: 9531303.
- 84. Jebbari H, Roberts CW, Ferguson DJ, Bluethmmann H, Alexander J. 1998. A protective role for IL-6 during early infection with *Toxoplasma gondii*. Parasite Immunol 20:231-239. doi:10.1046/j.1365-3024.1998.00152.x.
- 85. Scharton-Kersten TM, Yap G, Magram J, Sher A. 1997. Inducible nitric oxide is essential for host control of persistent but not acute infection with the intracellular pathogen *Toxoplasma gondii*. J Exp Med 185:1261-1273. doi:10.1084/jem.185.7.1261.
- 86. Alexander J, Scharton-Kersten TM, Yap G, Roberts CW, Liew FY, Sher A. 1997. Mechanisms of innate resistance to *Toxoplasma gondii* infection. Phil Trans R Soc Lond B Biol Sci 352:1355-1359. doi:10.1098/rstb.1997.0120.
- 87. Denkers EY and Gazzinelli R. 1998. Regulation and function of T-cell-mediated immunity during *Toxoplasma gondii* infection. Clin Microbiol Rev 11:569-588. doi:10.1128/CMR.11.4.569.
- 88. Starec M, Sinet M, Kodym P, Rosina J, Fiserova A, Desforges B, Rouveix B. 1997. The effect of drugs on the mortality of mice inoculated with Friend leukaemia virus or *Toxoplasma gondii*. Physiol Res 46:107-111. PMID: 9727501.
- 89. Stahl W, Dias JA, Turek G, Kaneda Y. 1995a. Etiology of ovarian dysfunction in chronic murine toxoplasmosis. Parasitol Res 81:109-113. doi:10.1007/BF00931615.
- 90. Stahl W, Kaneda Y, Tanabe M, Kumar SA. 1995b. Uterine atrophy in chronic murine toxoplasmosis due to ovarian dysfunction. Parasitol Res 81:114-120. doi:10.1007/BF00931614.

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